Compensatory growth of *Phanerochaete velutina* mycelial systems grazed by *Folsomia candida* (Collembola)

Sam Bretherton, George M. Tordoff, T. Hefin Jones & Lynne Boddy

Cardiff School of Biosciences, Cardiff, Wales, UK

Correspondence: Lynne Boddy, Cardiff School of Biosciences, Cardiff CF10 3TL, UK. Tel.: +44 029 20874776; fax: +44 029 20874305; e-mail: BoddyL@cf.ac.uk

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Abstract

Phanerochaete velutina is a major agent of wood decomposition in temperate forests. It grows out of woody resources in search of other resources and is then vulnerable to grazing by invertebrates. The aim of this study was to determine how continuous grazing and grazing for only 2 days by different densities of collembola, Folsomia candida, affect mycelial development (radial extension, hyphal coverage and fractal dimension) of *P. velutina* growing across non-sterile soil. High density (80 collembola) continuous grazing resulted in different mycelial foraging patterns compared to controls and lower density (20 and 40 collembola) continuous grazing: radial extension rate was reduced from 8.4 mm day^{-1} (control) to $6.9 \,\mathrm{mm}\,\mathrm{day}^{-1}$ (80 collembola), hyphal coverage was reduced to 81% of controls and mass fractal dimension increased from 1.68 (control) to 1.72 (80 collembola). There was evidence of over-compensatory growth: when high density grazing ceased the new growth was considerably greater (38%) than in controls. Grazing also resulted in growth stimulation: at low density continuous grazing (20 collembola) hyphal coverage was 15.6% greater than in controls. The ecological implications of compensatory and stimulatory growth in fungal-invertebrate interactions are considered.

Introduction

Mycelia of many fungal species that decompose wood and leaf litter are able to grow out of one resource in search of others. In doing so they may cross territory that is inhospitable in terms of available nutrients and microclimate, encounter antagonistic microorganisms and be grazed upon by invertebrates, including collembola, flies, nematodes and, in some localities, termites (Maraun et al., 2003). Grazing on fungal mycelia can result in marked changes in mycelial morphology (Hedlund et al., 1991; Kampichler et al., 2004; Harold et al., 2005; Tordoff et al., 2006) and enzyme production (Hedlund et al., 1991; Dyer et al., 1992), and increases in fungal respiratory activity (Hanlon & Anderson, 1979; Hanlon, 1981; Visser et al., 1981; Bengtsson & Rundgren, 1983; Bengtsson et al., 1993). In some cases greater growth (termed compensatory or 'catch-up' growth) has occurred following grazing, e.g. with the zygomycete Mortierella isabellina and the ascomycetes Penicillium spinulosum and Verticillium bulbillosum (Bengtsson & Rundgren, 1983; Bengtsson et al., 1993). Compensatory growth is a well-studied phenomenon across a wide range of macroorganisms, including herbivores grazing on plants and

vertebrates whose initial growth has been slow, for example due to suboptimal time of birth, poor prenatal or early postnatal nutrition (Trlica & Rittenhouse 1993; Agrawal, 2000; Metcalfe & Monaghan, 2001). Essentially, the organism experiences retarded growth and then enters a phase of accelerated growth once conditions improve. The present study investigates possible compensatory growth of *Phanerochaete velutina* following removal of grazing pressure.

There have been a great many studies on the effects of grazing on microfungi and mycorrhizal-formers (e.g. Finlay, 1985; McGonigle, 1995; Gange, 2000; Maraun *et al.*, 2003), but until recently few studies have centred on saprotrophic cord-forming basidiomycetes (Dyer *et al.*, 1992; Kampichler *et al.*, 2004; Harold *et al.*, 2005; Tordoff *et al.*, 2006). This is surprising considering the crucial central role that these fungi play in decomposition processes and nutrient cycling (Boddy, 1993; Boddy & Watkinson, 1994). Existing studies on saprotrophic basidiomycetes show that continuous collembola grazing can result in dramatic changes to mycelial morphology and foraging patterns, and the extent of these changes depends upon species of fungi, inoculum resource status, grazing intensity (density) and collembola species (Kampichler *et al.*, 2004; Harold *et al.*, 2005; Tordoff *et al.*, 2005; Tordoff *et al.*, 2005;

2006). Further, selective grazing can influence the vertical distribution of fungi in soil (Newell 1984a, b). In the field, grazing pressure will vary in time and space, but little if anything is known about the behaviour of mycelia following removal of grazing pressure (i.e. resilience). This paper investigates: (1) the density dependent effects of grazing by the collembola Folsomia candida on mycelia of the cordforming basidiomycete P. velutina in soil microcosms in the laboratory; and (2) recovery of the mycelium when grazing ceases (i.e. when collembola are removed). Previous studies (Kampichler et al., 2004; Tordoff et al., 2006) suggest that heavy grazing will have the greatest effects in terms of reduction in hyphal coverage and changes to mycelial morphology (fractal dimension). It is hypothesized that high intensity grazing will reduce radial extension rate and hyphal coverage, and that when grazing stops, heavily grazed mycelia will take longer to recover than lightly grazed mycelia.

Materials and methods

Fungal isolate and inoculum preparation

Phanerochaete velutina (Cardiff University culture collection; Dowson *et al.*, 1986) was subcultured on 2% malt extract agar (MEA: 20 g L^{-1} Munton & Fison spray malt, 15 g L^{-1} Lab M agar no. 2). Petri dishes (14 cm diameter) were colonized with *P. velutina* for 12 days before 15 beech (*Fagus sylvatica*) wood blocks ($2 \times 2 \times 1$ cm) were added, and incubated for 3 months in the dark at 20 °C. The wood blocks had been cut from a freshly-felled tree and stored at -18 °C until required. Prior to use, the blocks were soaked overnight in deionized water to defrost and autoclaved at 121 °C for 20 min in sealed autoclave bags, three times at 24 h intervals.

Culturing and extraction of F. candida

Folsomia candida (supplied by Centre for Ecology and Hydrology, Lancaster, UK) were reared on a 90% plaster of Paris (Minerva Dental Ltd, Cardiff, UK): 10% charcoal (Sigma, UK) substrate in 0.9 L plastic containers, at room temperature. Culture boxes had holes in the lid to provide aeration and collembola were fed weekly with dried baker's yeast (*Saccharomyces cerevisiae*; Spice of Life, Cardiff, UK) and the substrate kept moist with deionized water.

Collembola were extracted from cultures using a series of stacked metal sieves (Nickel-Electro Ltd., Weston-super-Mare, UK) of progressively smaller pore size. Those of the required size (250–400 μ m) were transferred to new culture boxes and starved for 24 h prior to use in the experiments. Individuals were collected for introduction into each microcosm using an electrical aspirator (pooter).

Preparation of soil microcosms

Soil (0–20 cm depth) was collected from mixed deciduous woodland in the Coed Beddick Inclosure, Tintern, UK. Wood and leaf litter were removed and the soil was sieved through a 10 mm mesh. The soil was air-dried for 14 days, sieved firstly through ≤ 4 mm mesh then through ≤ 2 mm mesh, and frozen (-18 °C) for 24 h to defaunate. Soil was then thoroughly mixed with an appropriate volume of deionized water to achieve a matric potential of -0.012 MPa. Wet soil (200 g) was added to 24×24 cm lidded bioassay trays (Nunc-Gibco, Paisley, UK), and compacted evenly and smoothly to approximately 5 mm depth.

Soil trays (n = 70) were inoculated by positioning a colonized wood block, which had been first scraped free of adhering agar and mycelium using a scalpel, at the centre of each tray. The weight of each soil tray, including inoculum, was recorded; every 7 days, trays were re-weighed and remoistened by evenly spraying a mist of deionized water onto uncolonized regions of soil. Trays were kept individually in sealed polythene bags (to reduce moisture loss and prevent collembola escaping) and stacked randomly in the dark at 20 ± 1 °C.

Experimental design

Seven treatments with 10 replicates each were carried out, each treatment assigned randomly amongst the 70 trays: ungrazed control; 20, 40 or 80 collembola added at 10 days (by which time mycelia were at least 8 cm diameter) and allowed to continue grazing for the duration of the experiment; 20, 40 or 80 collembola added at 10 days and then removed 2 days later. Collembola were removed using an electrical aspirator taking care not to damage the mycelium. Although 80 collembola per tray represents only 1566 m^{-2} soil, which is less than usual field densities $(10^4-10^5 \text{ m}^{-2})$ (Petersen & Luxton, 1982), this is appropriate since the collembola are restricted to two dimensions in the microcosms, whereas the field figures quoted are for soil cores of three dimensions.

Image capture and analysis

Digital images of experimental systems were captured immediately prior to collembola addition and then after 2, 3, 4, 5, 7, 9 and 12 days, with a Sony Cyber Shot digital still camera positioned 60 cm above the microcosm, with natural lighting. Saved JPEG images were processed using IMAGEJ 1.33u software (National Institute of Health, USA) in a Microsoft Windows XP environment. Tray margins and wood inocula were electronically removed from all digital images prior to conversion to greyscale (8-bit). Images were then subject to manual thresholding: any pixels with a grey value less than the threshold were converted to black to represent soil, and pixels with values greater than the threshold were converted to white, representing mycelium. Radial extension measurements were obtained by electronically measuring eight lines radiating at 45° angles from the position of the wood block inocula to the mycelial margin. The pixel length of a 22.6 cm line (the internal length of the side of a bioassay tray) was used for calibration. Determination of radial extent ceased when mycelia reached the edges of the trays. Hyphal coverage was determined as the number of white pixels in a binary image, converted by IMAGEJ to cm². Fractal dimension provides a good measure of space filling. Phanerochaete velutina is approximately mass fractal (i.e. the whole mycelium, not just the border, is fractal; Boddy et al., 1999) and hence only the mass fractal dimension (D_{BM}) was determined in IMAGEJ by the box-counting method, using the Fractional Dimension and Lacunarity Plug-in.

As well as analysis of each entire image, images at 12 days after collembola addition were divided electronically into two parts – the area bounded by the mycelial front at 2 days (i.e. when collembola were removed from 30 trays), and that between the line of this front and the mycelial front at 12 days (Fig. 1). Hyphal coverage was determined for the outer zone.

Statistical analyses

Radial extension, hyphal coverage and D_{BM} were analyzed by one-way Repeated Measures Analysis of Variance



Fig. 1. A binary image of a mycelial system of *Phanerochaete velutina* grazed for 12 days. Line indicates the position of the mycelial front after 2 days of grazing. Hyphal coverage was determined for the entire system and separately for the outer zone that had developed between 2 and 12 days.

(RMANOVA; SPSS, release 12), using grazing treatments as the main effect and time as a sub-factor. Data were normally distributed (Kolmogorov-Smirnov test), had equal variance (Levene's test) and displayed sphericity (Mauchly's test of sphericity), and therefore met assumptions for RMANOVA. Significant results were explored further using the Tukey's pairwise comparison to determine significant differences between means at individual time points. Data are presented with standard error of the mean.

Results

Grazing for 2 days caused dramatic differences in mycelial morphology, especially in the high density (80 collembola) grazing treatment, where mycelium was denser in central regions and there was prolific fanning at the mycelial margin (Fig. 2). Grazing was largely at the mycelial margin and, to a lesser extent, close to the inoculum. When grazing pressure was removed from these systems, subsequent growth looked similar to the control, but was sometimes quantitatively different (see below).

Radial extension

Grazing did not significantly (P > 0.05) affect extension rate relative to ungrazed systems (Fig. 3). The extension rate of mycelia continuously grazed by 40 or 80 collembola was less than in those that were only grazed for 2 days, though not significantly (P > 0.05). The extension rate of systems grazed continuously by 20 collembola was, however, significantly ($P \le 0.05$) faster than in systems only grazed for 2 days.

Hyphal coverage

In all treatments, hyphal coverage increased linearly until 5 days after collembola addition (when sides of trays were reached) and then did not rise (Fig. 4a–c). By 12 days after collembola addition there was, however, an increase in hyphal coverage in systems continuously grazed by 20 collembola and systems which had been grazed by 80 collembola for 2 days. Hyphal coverage was similar for all treatments except for that continuously grazed for 80 days, which was significantly (P=0.02) lower.

Hyphal coverage in the area which developed after 2 days was significantly ($P \le 0.05$) less in systems continuously grazed by 80 collembola than in all other systems (Fig. 5). In systems which had been grazed for 2 days by 80 collembola or continuously grazed by 20 collembola, hyphal coverage was significantly ($P \le 0.05$) greater than in ungrazed controls. Other grazing treatments were not significantly (P > 0.05) different from ungrazed controls.



Fig. 2. Digital images of mycelial systems of *Phanerochaete velutina* (in 24×24 cm trays of compressed non-sterile soil) that had been ungrazed (a, b), continuously grazed (C: c, d, g, h, k, l) or grazed for 2 days then grazing pressure removed (R: e, f, i, j, m, n). Images captured 2 days (a, c, e, g, i, k, m) and 12 days (b, d, f, h, j, l, n) after adding collembola (time of image capture indicated at head of column of images). Collembola were added at different densities (indicated in left margin): 0 (a, b), 20 (c–f), 40 (g–j) or 80 (k–n) per tray.



Fig. 3. Extension rate of *Phanerochaete velutina* mycelium subject to different grazing intensity (0, 20, 40, 80 collembola per tray). Grazing was for only 2 days in half of the treatments (indicated by R) or continued (indicated by C) for the duration of the experiment (12 days). Error bars are the standard error of the mean. Bars with the same letter are not significantly (P > 0.05) different.

Mass fractal dimension

 D_{BM} was at a maximum at about 5 days and then decreased (Fig. 6). Systems that were only grazed for 2 days were not significantly (P > 0.05) different from ungrazed controls. Continuously grazed systems had consistently higher D_{BM} than other systems, though this was not significantly (P > 0.05) different.

Discussion

High intensity grazing (at least in the context of this experiment) dramatically affected fractal dimension, hyphal coverage and overall morphology, but not the extension rate, of mycelium of the cord-forming basidiomycete *P. velutina*, only partly confirming the hypothesis that high intensity grazing will reduce radial extension rate and hyphal coverage. This study did not support earlier work which found that the extension rate of *P. velutina* was dependent on the density of grazing *F. candida*, and that there was no effect on hyphal coverage (Tordoff *et al.*, 2006). This may have been due to the use of smaller trays in the earlier study. Grazing intensity was also important in systems of *Hypholoma fasciculare* and *Resinicium bicolor* (Tordoff *et al.*, 2006).

The second hypothesis, that heavily grazed systems will take longer to recover than those grazed more lightly, was rejected. Mycelial systems rapidly recovered from 2 days high intensity grazing, and hyphal coverage, in the region of mycelium that developed after removal of grazers, was greater in the high intensity grazing (80 collembola) systems than in controls. Equally significantly, systems subject to low intensity (20 collembola) continuous grazing exhibited



Fig. 4. Change in *Phanerochaete velutina* hyphal coverage with time in mycelial systems grazed by 20 (a), 40 (b) or 80 (c) collembola compared with ungrazed systems. \Box ungrazed; \blacktriangle continuously grazed; \bigcirc grazed for 2 days then grazing ceased. Repeated measures ANOVA revealed a significant (F_{36,378}=2.871, *P* < 0.001) time × treatment interaction, but no overall treatment effect (i.e. irrespective of time) (F_{6,63}=1.897, *P*=0.095). *Significant difference (*P*=0.002) between continuously grazed and grazed for 2 days.

greater hyphal coverage (compared to ungrazed controls) in the mycelial area that developed following addition of collembola, i.e. compensatory growth. Often compensatory growth is incomplete, with deprived organisms being unable to reach the size of unaffected controls but over-compensation, such as that in the present study, does sometimes occur, with deprived/stressed individuals becoming larger Hyphal coverage (cm²)

0

0

20 R



1.76

1.74

1 72

1.70

1.68

1.66

1.64

1.76

1.74

1.72

1.70

1.68

1.66

1.64

1.76

1.74

1.72

1.70

1.68

1.66

1.64

0

2

4

Mass Fractal Dimension (D_{BM})

Fig. 5. Hyphal coverage of *Phanerochaete velutina* mycelium subject to different grazing intensity (0, 20, 40, 80 collembola per tray) in the zone that developed after grazing (see Fig. 1). Grazing was for only 2 days in half of the treatments (indicated by R) or continued (indicated by C) for 12 days until the end of the experiment. Error bars are the standard error of the mean. Bars with the same letter are not significantly (P > 0.05) different

40 R

Grazing treatment

40 C

80 R

80 C

20 C

than controls (Detling & Painter, 1983; Hayward et al., 1997). Determining a mechanism by which a heavily grazed organism will eventually grow larger than ungrazed ones is difficult (McNaughton, 1983; Paige, 1992; Trumble et al., 1993; Callaway et al., 2001). For plants it has been suggested that removal of apical dominance and/or growth promoters produced by the herbivores may be a means by which plants benefit from grazing, producing, for example, multiple flowering stalks (Paige & Whitham, 1987; Paige, 1992; Dyer et al., 1995; Tiffin, 2000). Similar increased branching occurred in the present study. A further compensatory mechanism in plants results from increased photosynthesis, possibly due to increased light penetration in grazed areas resulting in decreased 'competition' by leaves for light, and also decreased competition for water and nutrients (McNaughton, 1983; Dyer et al., 1995). Decreased competition for water and nutrients may also apply to the grazed mycelia in the microcosms.

Stimulatory effects of grazing have never been reported for saprotrophic basidiomycetes (Kampichler *et al.*, 2004; Harold *et al.*, 2005; Tordoff *et al.*, 2006). Indeed, in previous studies continuous grazing has resulted in decreases in hyphal coverage and radial extension rates rather than the increased hyphal coverage observed with the low density grazing (20 collembola) in the present study. Why growth of *P. velutina* should be stimulated in this study and not in the previous study with similar grazing pressure (Tordoff *et al.*, 2006) is not clear, but it may relate to differences in the size of the soil trays used, which were smaller previously.

Compensatory growth of plants often carries a cost. For example, storage reserves may be utilized, and growth of replacement tissues may be at the expense of other growth



Fig. 6. Change in *Phenerochaete velutina* mass fractal dimension (D_{BM}) with time in mycelial systems grazed by 20 (a), 40 (b) or 80 (c) collembola compared with ungrazed. \Box ungrazed; \blacktriangle continuously grazed; \bullet grazed for 2 days then grazing ceased. Repeated measures ANOVA revealed no significant (F_{36,378}=0.851, *P*=0.716) time × treatment interaction, but a significant (F_{6,63}=2.884, *P*=0.015) overall treatment effect (i.e. irrespective of time).

6

Days after collembola addition

8

10

12

and metabolic centres, or of reproduction (Tiffin, 2000; Trumble *et al.*, 2003; Pratt *et al.*, 2005). Such costs can be paid over a range of time scales (Metcalfe & Monaghan, 2001). Mycelia could operate a trade-off between extension rate of mycelia and hyphal coverage, though there is no evidence of that in this study. Here the trade-off for increased coverage is probably the more rapid use of the

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wood inoculum reducing the length of time that the fungus could utilize the resource.

A variety of evolutionary responses to herbivory have been noted for plants, including resistance, mutualism, over-compensation, phenological escape and tolerance (Agrawal, 2000). Of these, resistance, phenological escape and tolerance are all defensive mechanisms that increase fitness by reducing grazing. With fungi, resistance to grazing could be effected by the production of allelopathic chemicals on the surface of individual hyphae and cords, or as volatiles or secondary metabolites produced when hyphae are damaged. Mycelial cords of some basidiomycetes have crystals, especially of calcium oxalate, on their surface, and these may inhibit invertebrate grazing (Connolly & Jellison, 1995). Phenological escape (i.e. plants not being available when herbivores are most active) may have an analogue with fungi: although basidiomycete cord-formers produce extensive perennial systems (Boddy, 1993; Cairney, 2005), they are dynamic with new outgrowth from extending foraging fronts and from newly colonized organic resources (Cairney, 2005). There is evidence that during winter months, mycelium of P. velutina not interconnecting with resources dies back and new growth from a resource only then occurs when conditions improve (L. Boddy, R. G. Bolton & S. H. Abdallah, pers. commun.). If renewed outgrowth were timed to coincide with times of reduced grazer activity, this might be considered phenological escape. Tolerance implies little or no reduction in fitness as a result of grazing; in the present study, hyphal coverage of systems grazed by 40 collembola was not significantly different from that of ungrazed controls.

Grazing resulting in stimulation/over-compensation of mycelial growth, as seen in the present study, or collembola recycling nutrients through production of faeces, and removal of competing microbes from the soil surface may be regarded as mutualism. Increased mycelial growth, however, should only be regarded as beneficial if more resources are obtained to compensate for the more rapid use of the organic resource from which the fungus is obtaining its carbon and mineral nutrients for mycelial extension. Since compensatory growth is presumably largely fed by decomposition of organic resources interconnected to the mycelium in soil, it is likely that mycelia will compensate to different extents depending on the quality and quantity of resources available, and the relative size of the mycelium. Similarly, in plants utilization of stored reserves is likely to be an important mechanism of tolerance to grazing, and some studies have shown a positive correlation between root-shoot ratios and regrowth following defoliation (Tiffin, 2000).

Having found evidence of dramatic density dependent grazing effects in fungi, including compensatory and stimulatory growth, we now need to know (1) how widespread this phenomenon is in fungi, (2) whether other types of mechanical damage have similar effects to invertebrate grazing, and (3) whether grazing and mechanical damage affect mycelia in the field to the same extent as in the

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References

laboratory.

- Agrawal AA (2000) Overcompensation of plants in response to herbivory and the by-product benefits of mutualism. *Trends Pl Sci* 5: 309–313.
- Bengtsson G & Rundgren S (1983) Respiration and growth of a fungus, *Mortierella isabellina*, in response to grazing by *Onychiurus armatus* (Collembola). *Soil Biol Biochem* 15: 469–473.
- Bengtsson G, Hedlund K & Rundgren S (1993) Patchiness and compensatory growth in a fungus-collembola system. *Oecologia* 93: 296–302.
- Boddy L (1993) Saprotrophic cord-forming fungi: warfare and other ecological aspects. *Mycol Res* 97: 641–655.
- Boddy L (1999) Saprotrophic cord-forming fungi: meeting the challenge of heterogeneous environments. *Mycologia* 91: 13–32.
- Boddy L & Watkinson SC (1994) Wood decomposition, higher fungi, and their role in nutrient redistribution. *Can J Bot* 73 (Suppl. 1): S1377–1383.
- Boddy L, Wells JM, Culshaw C & Donnelly DP (1999) Fractal analysis in studies of mycelium in soil. *Geoderma* 88: 301–328.
- Cairney JWG (2005) Basidiomycete mycelia in forest soils: dimensions, dynamics and roles in nutrient distribution. *Mycol Res* **109**: 7–20.
- Callaway RM, Newingham B, Zabinski CA & Mahall BE (2001) Compensatory growth and competitive ability of an invasive weed are enhanced by soil fungi and native neighbours. *Ecol Lett* **4**: 429–433.
- Connolly J & Jellison J (1995) Calcium translocation, calcium oxalate accumulation, and hyphal sheath morphology in the white rot fungus *Resinicium bicolor. Can J Bot* **73**: 927–936.
- Detling JK & Painter EL (1983) Defoliation responses of Western wheatgrass populations with diverse histories of prairie dog grazing. *Oecologia* **57**: 65–71.
- Dowson CG, Rayner ADM & Boddy L (1986) Outgrowth patterns of mycelial cord-forming basidiomycetes from and between woody resource units in soil. *J Gen Microbiol* **132**: 203–211.
- Dyer HC, Boddy L & Preston-Meek CM (1992) Effect of the nematode *Panagrellus redivivus* on growth and enzyme production by *Phanerochaete velutina* and *Stereum hirsutum*. *Mycol Res* **96**: 1019–1028.

Dyer MI, Moon AM, Brown MR & Crossley DA Jr (1995) Grasshopper crop and midgut extract effects on plants: an example of reward feedback. *Proc Natl Acad Sci USA* **92**: 5475–5478.

Finlay RD (1985) Interactions between soil micro-arthropods and endomycorrhizal associations of higher plants. *Ecological Interactions in Soil. Plants, Microbes and Soil Animals* (Fitter AH, Atkinson D, Read DJ & Usher MB, eds), pp. 319–331. Blackwell Scientific, Oxford, UK.

Gange A (2000) Arbuscular mycorrhizal fungi, collembola and plant growth. *Trends Ecol Evol* **15**: 369–372.

Hanlon RDG (1981) Influence of grazing by collembola on the activity of senescent fungal colonies grown on media of different nutrient concentration. *Oikos* **36**: 362–367.

Hanlon RDG & Anderson JM (1979) The effects of collembola grazing on microbial activity in decomposing leaf litter. *Oecologia* **38**: 93–99.

Harold S, Tordoff GM, Jones TH & Boddy L (2005) Mycelial responses of *Hypholoma fasciculare* to collembola grazing: effect of inoculum age, nutrient status and resource quality. *Mycol Res* **109**: 927–935.

Hayward RS, Noltie DB & Wang N (1997) Use of compensatory growth to double hybrid sunfish growth rates. *Trans Am Fish Soc* **126**: 316–322.

Hedlund K, Boddy L & Preston CM (1991) Mycelial responses of the soil fungus *Mortierella isabellina* to grazing by *Onychiurus armatus* (Collembola). *Soil Biol Biochem* 23: 361–366.

Kampichler C, Rolschewski J, Donnelly DP & Boddy L (2004) Collembolan grazing affects the growth strategy of the cordforming fungus *Hypholoma fasciculare*. Soil Biol Biochem 36: 591–599.

Maraun M, Martens H, Migge S, Theenhaus A & Scheu S (2003) Adding to 'the enigma of soil animal diversity': fungal feeders and saprophagous soil invertebrates prefer similar food substrates. *Eur J Soil Biol* **39**: 85–95.

McGonigle TP (1995) The significance of grazing on fungi in nutrient cycling. *Can J Bot* **73** (Suppl. 1): S1370–S1376.

McNaughton SJ (1983) Compensatory plant growth as a response to herbivory. *Oikos* **40**: 329–336.

Metcalfe NB & Monaghan P (2001) Compensation for a bad start: grow now, pay later. *Trends Ecol Evol* **16**: 254–260.

Newell K (1984a) Interaction between two decomposer Basidiomycetes and a collembolan under Sitka spruce: distribution, abundance and selective grazing. *Soil Biol Biochem* 16: 227–233.

Newell K (1984b) Interaction between two decomposer Basidiomycetes and a collembolan under Sitka spruce: grazing and its potential effects on fungal distribution and litter decomposition. *Soil Biol Biochem* **16**: 235–239.

Paige KN (1992) Overcompensation in response to mammalian herbivory: from mutualistic to antagonistic interactions. *Ecology* **73**: 2076–2085.

Paige KN & Whitham TG (1987) Overcompensation in response to herbivory: the advantage of being eaten. Am Nat 129: 407–416.

Petersen H & Luxton M (1982) A comparative analysis of soil fauna populations and their role in decomposition processes. *Oikos* **39**: 287–388.

Pratt PD, Rayamajhi MB, Van TK, Center TD & Tipping PW (2005) Herbivory alters resource allocation and compensation in the invasive tree *Melaleuca quinquenervia*. *Ecol Entomol* 30: 316–326.

Tiffin P (2000) Mechanisms of tolerance to herbivore damage: what do we know? *Evol Ecol* 14: 523–536.

Tordoff GM, Jones TH & Boddy L (2006) Grazing by *Folsomia candida* (Collembola) affects mycelial morphology of the cord-forming basidiomycetes *Hypholoma fasciculare*, *Phanerochaete velutina* and *Resinicium bicolor*. *Mycol Res* **110**: 335–345.

Trlica MJ & Rittenhouse LR (1993) Grazing and plant performance. *Ecol Appl* **3**: 21–23.

Trumble JT, Kolodny-Hirsch DM & Ting IP (1993) Plant compensation for herbivory. *Ann Rev Ent* **38**: 93–119.

Visser S, Whittaker JB & Parkinson D (1981) Effects of collembolan grazing on nutrient release and respiration of a leaf litter inhabiting fungus. *Soil Biol Biochem* 13: 215–218.